

High throughput sequencing of fungal communities

Ari Jumpponen

Division of Biology

Ecological Genomics Institute

Kansas State University

Ken Jones

Environmental Health Science

University of Georgia

Outline

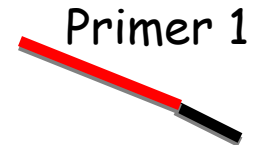
- High throughput sequencing?
 - DNA tags as sample IDs
 - 454 pyrosequencing platform
- Sequencing of regions on 454 platform
- Trade offs - depth vs. sample numbers
 - Trade offs - costs

High throughput sequencing

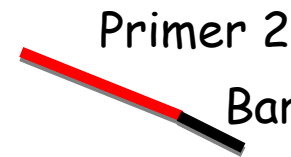
- Sequencing of a large number of templates from heterogeneous samples
 - Clone libraries and Sanger sequencing
 - Automated colony picking
 - 96- or 384-well formats for extension/sequencing
 - Direct sequencing on 454 pyrosequencing platform
 - Incorporation of 454 endemic priming sites
 - emPCR signal amplification and massively parallel sequencing
- Both can use DNA tagging (primer barcoding for sample ID)

Primer tagging - DNA barcoding

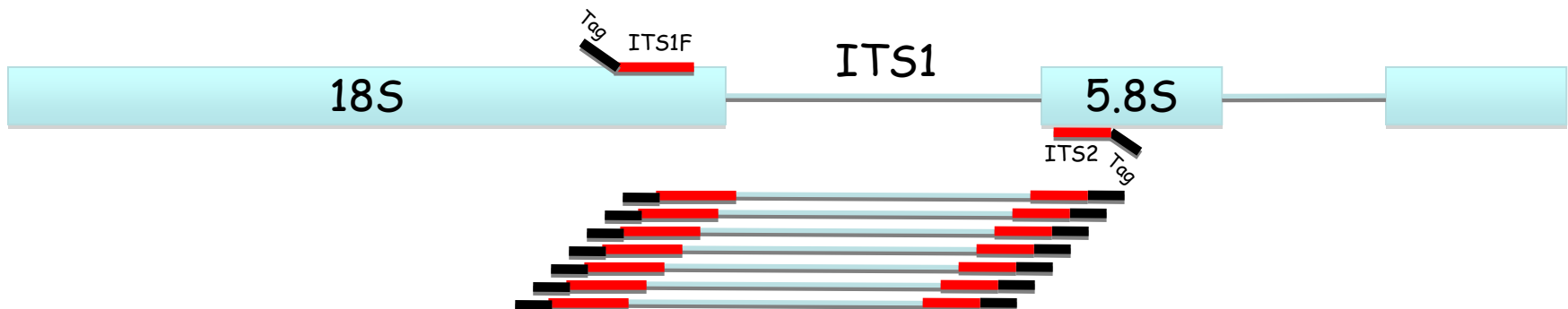
- Use a user-determined identifier DNA-tag
- Tagging both primers allows non-directional sequencing with vector-residing primers
- The tags can be added by ligation into A-overhang, directly in PCR or in a nested PCR reaction



Bar-code (DNA Tag)

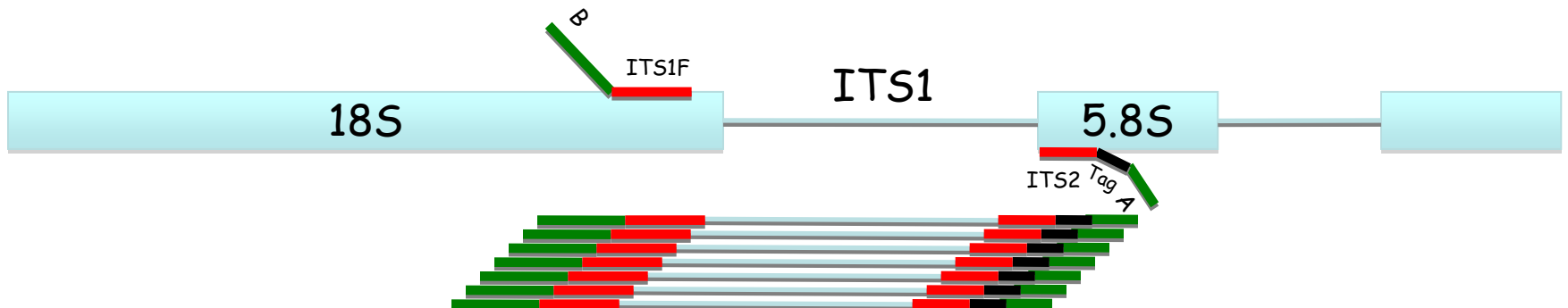
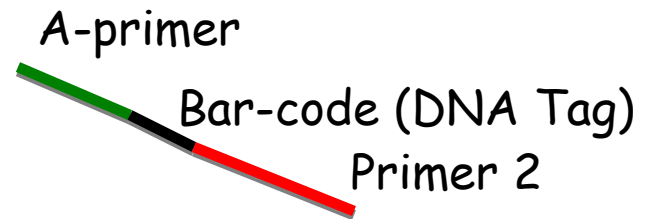
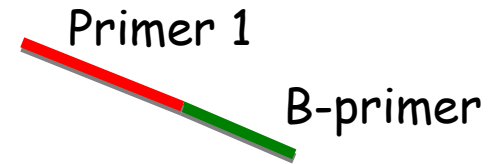


Bar-code (DNA Tag)



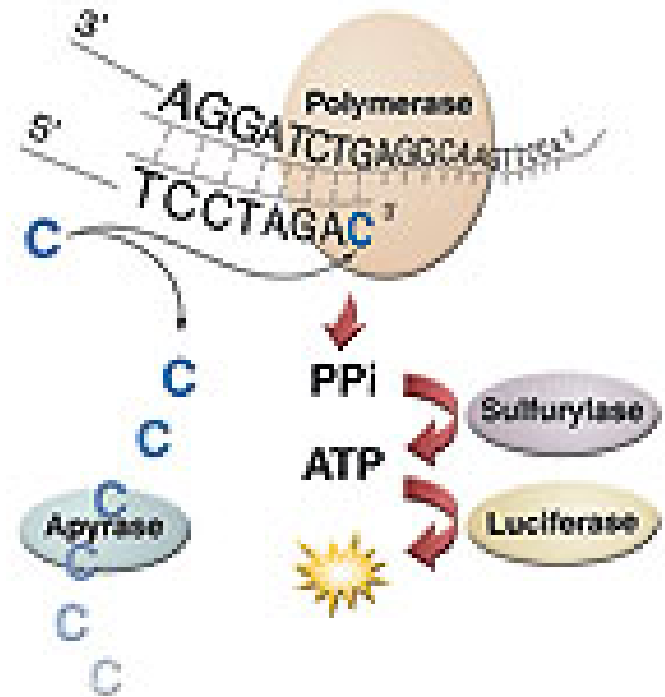
Amplification for 454 pyrosequencing

- Since the sequencing is done with a platform endemic primer, one barcoded primer necessary



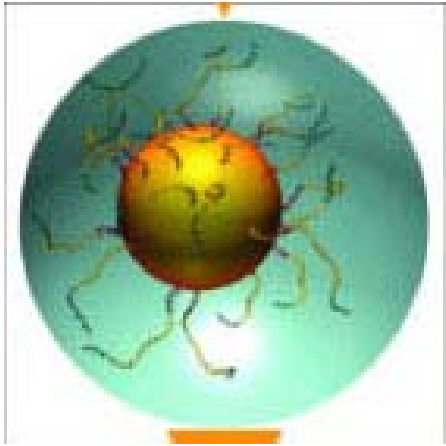
What is pyrosequencing?

- Developed by Ronaghi and Nyrén (Analytical Biochemistry 1996 and Science 1998) for BioTag, licenced now to 454 LifeSciences (bought by Roche 2007)
- In brief, DNA polymerase incorporation of phosphorylated dNTP's emits light.
- Sequencing way faster than with the chain termination method (Sanger).
- Huge throughput: 100M nucleotides in 7 hour run.

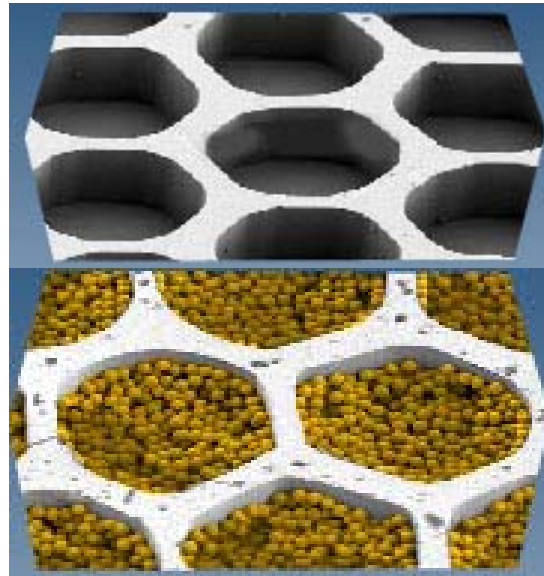


Massively Parallel 454 Sequencing

Emulsion PCR (emPCR) amplification of the PCR products in microreactor bubbles



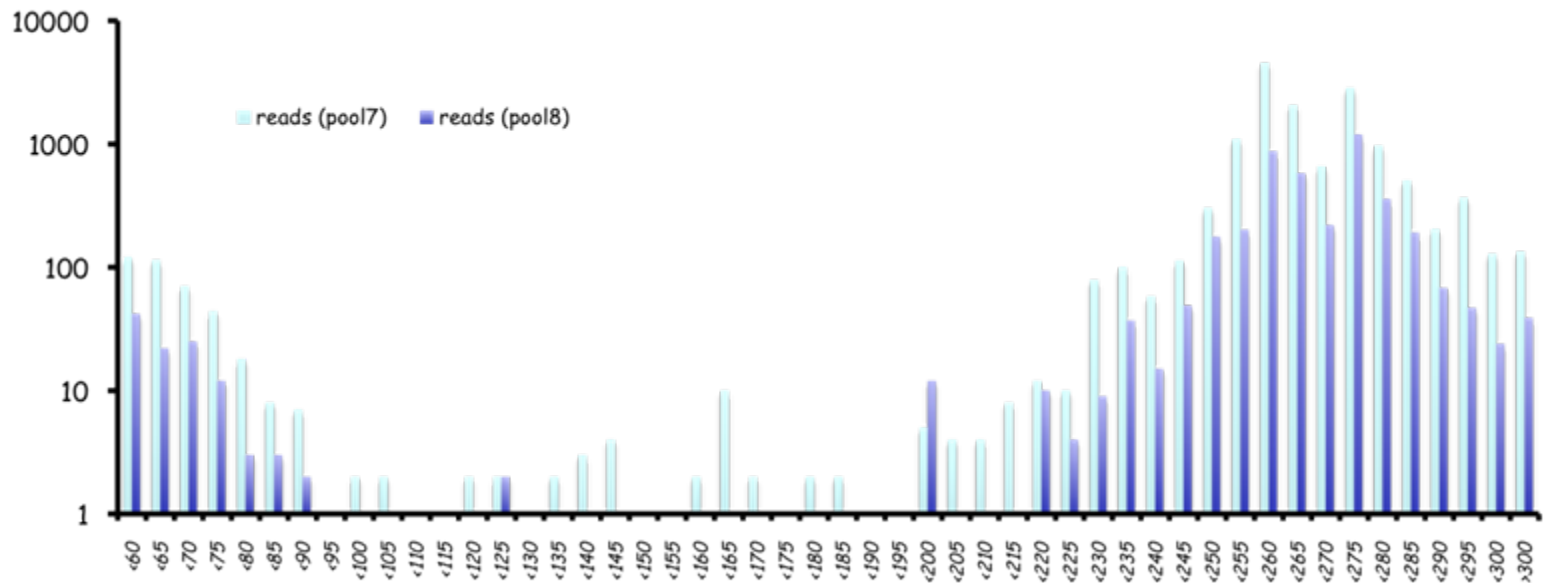
Dispense beads to picotiter plates and fill packing beads



Sequence via pyrosequencing

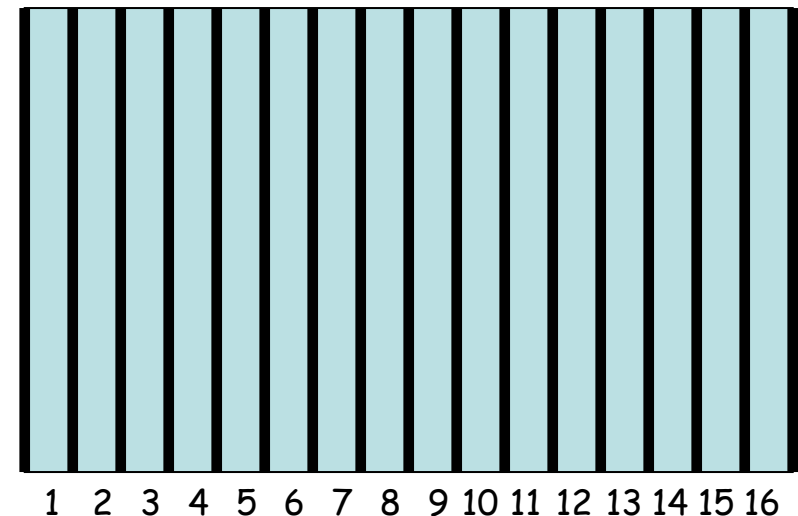
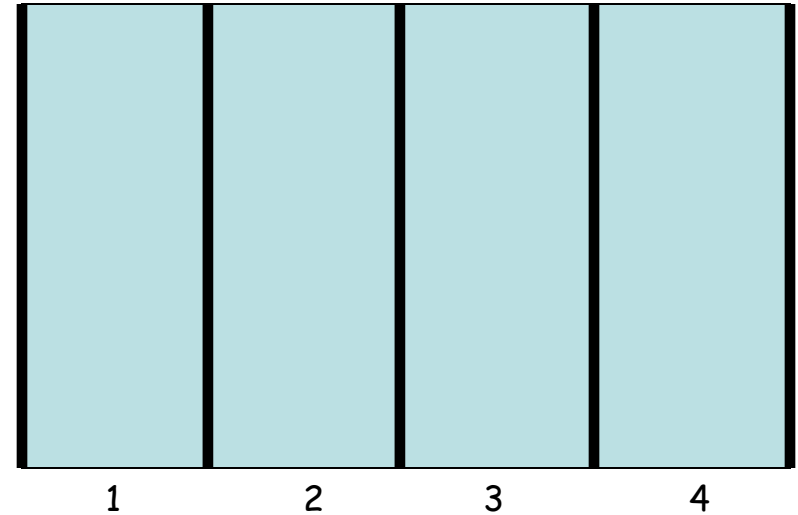
454 yields

- Up to 250,000 reads, 265bp in length
- Sequence quality control
 - ID Primer
 - ID Barcode
 - Ambiguous bases



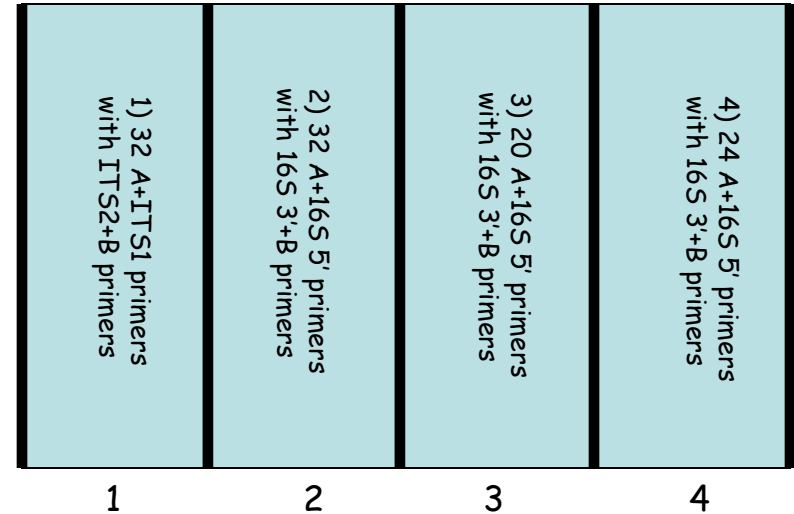
454 sequencing in regions

- RXN blocking
 - Each 454 rxn can be divided to 4 or 16 blocks
 - Loss of 10% (4 blocks) or 15-20% (16 blocks) of the throughput
 - Independent amplicon/data generation
- Combination of 16 regions and 64 tagged primers allows sequencing of 1,064 templates at ~150 reads per sample



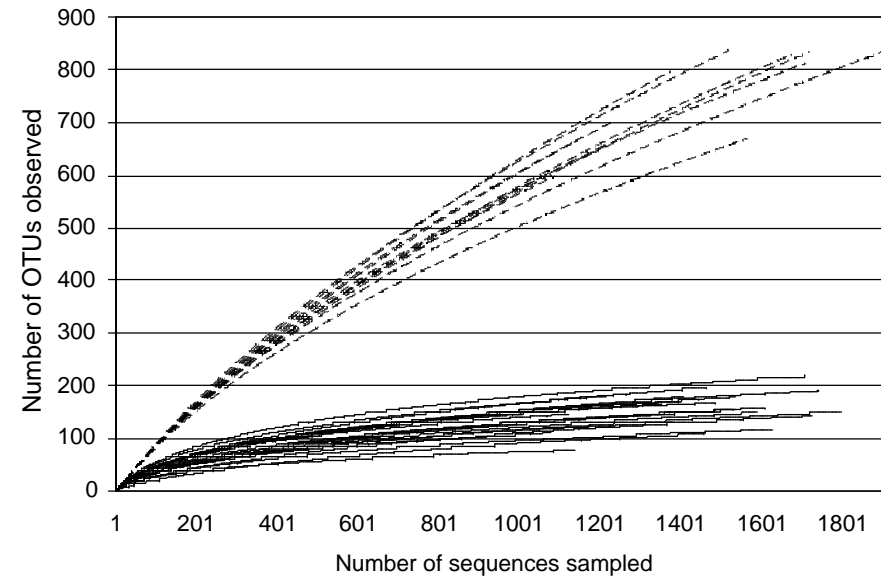
Example of region separation

- 4 experiments in 4 block + barcoding
 - 1) 32 barcoded (5bp barcodes) samples to study microeukaryon responses to rainfall manipulation and warming.
 - 2) 32 barcoded (5bp barcodes) samples to study bacterial responses to rainfall manipulation and warming
 - 3) 20 barcoded (5bp barcodes) samples *Peromyscus* gut microbes in contaminated and control sites
 - 4) 24 barcoded samples to study bacterial responses to N deposition
 - = 108 samples for a total of $\geq 160,000$ reads;
~1,500 reads per sample



Acquired Data - Numbers

- Rainfall Manipulation Plots (RaMPs)
- 1,200-1,800 reads per soil sample
- Eubacteria
 - 44,363 16S reads
 - 7,190 non-singletons (72% of the reads) + 12,439 singetons
 - Total 19,629 OTUs
- Eukarya
 - 41,512 ITS1 reads
 - 1,802 non-singletons (92% of the reads) + 3,264 singetons
 - Total 5,066 OTUs
- Eukarya nearly saturated, bacteria likely require 10x more sampling...

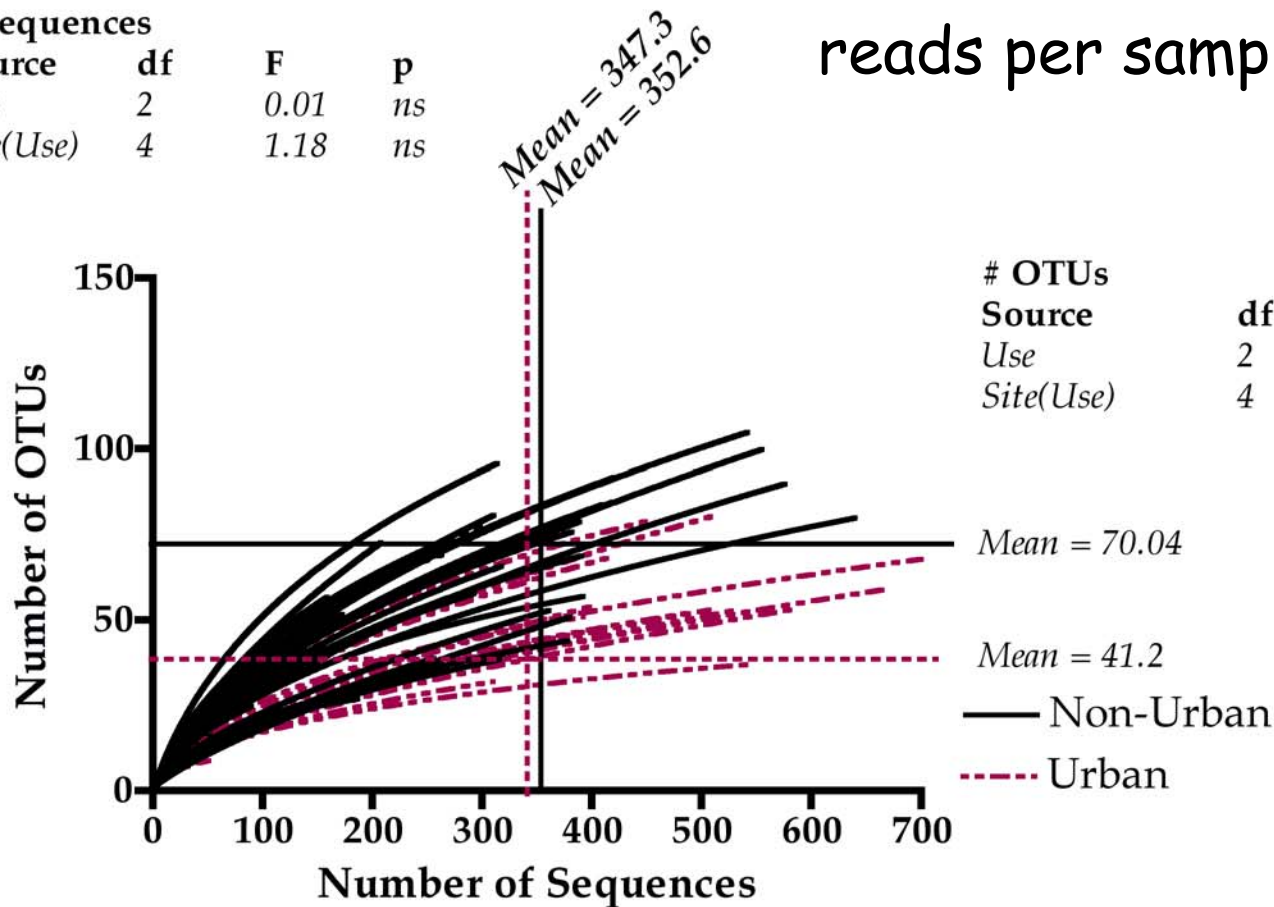


OTU richness per sample. Dashed line - bacteria; solid line - fungi.

Depth vs. number of samples

Foliar endophyte communities:
no saturation at 300-700
reads per sample

# Sequences			
Source	df	F	p
Use	2	0.01	ns
Site(Use)	4	1.18	ns



# OTUs			
Source	df	F	p
Use	2	13.08	<.0001
Site(Use)	4	1.40	ns

Mean = 70.04

Mean = 41.2

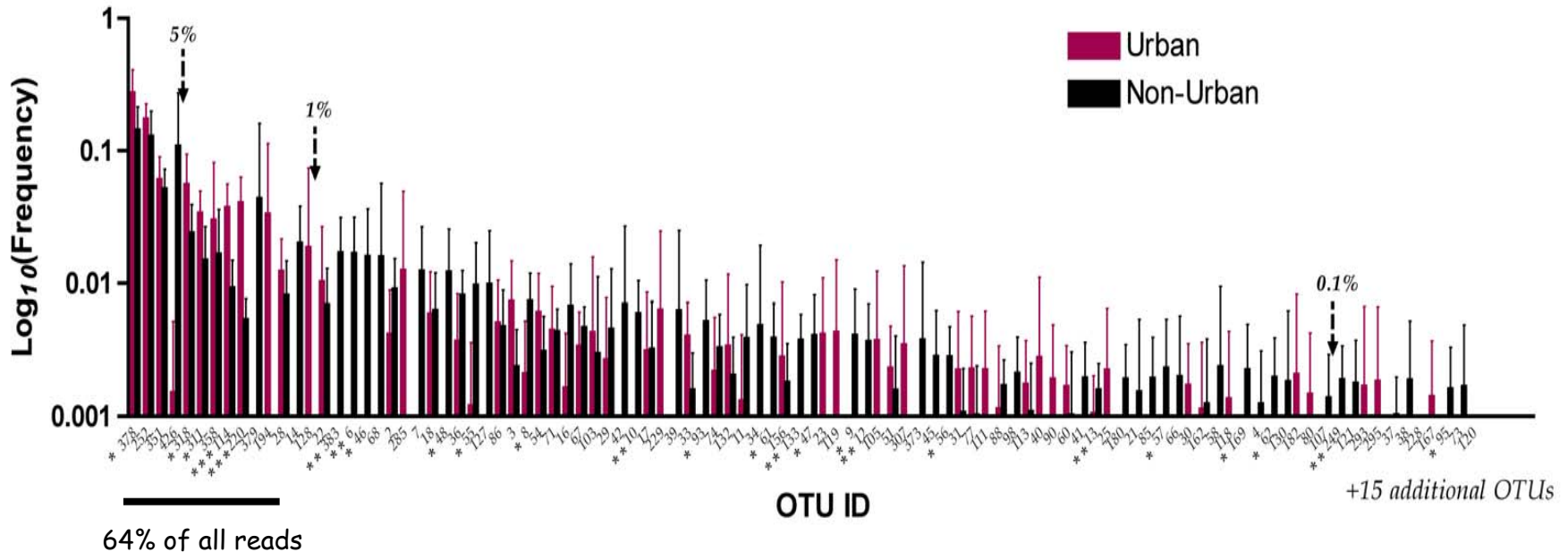
— Non-Urban

- - - Urban

Depth vs. number of samples

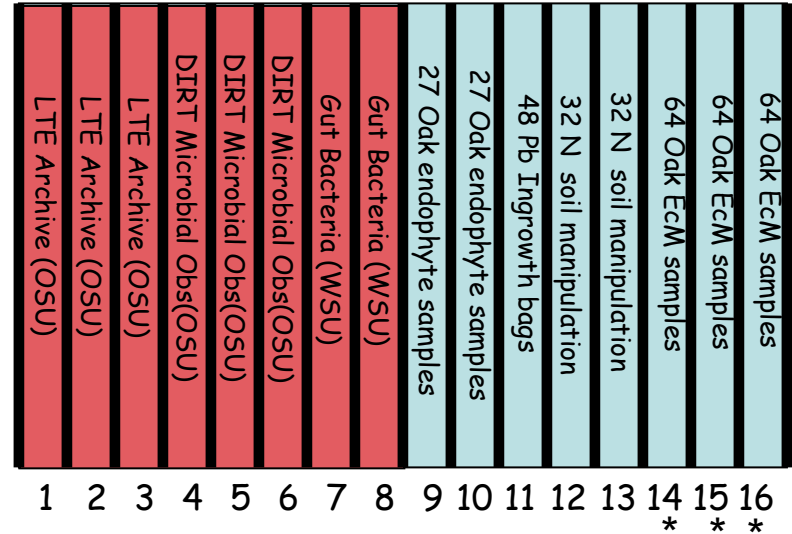
Foliar endophyte communities:

- 1) Top 10 most abundant OTUs comprised 64% of all reads
- 2) Only 14 most abundant OTUs $\geq 1\%$ of all reads
- 3) Only 4 most abundant OTUs $\geq 5\%$ of all reads



Costs - Urban Ectomycorrhizas

- 16 region RXN divided between 3 labs
- 6 microbial expts in 14 regions
 - 14-16) 64 barcoded EcM samples for each of 3 seasonal samples
 - 12-13) 2x32 barcoded LTER soil samples
 - 11) 48 barcoded fungal samples from Pb contaminated and control sites
 - 9-10) 54 barcoded samples for oak foliar endophytes
 - (7-8) *Peromyscus* gut microbiome)
 - 4-6) 20 soil archive samples
 - 1-3) 12 DIRT microbial observatory samples
 - = total of 385 samples for ~100,000 reads; average of ~260 reads per sample
- Total 454 cost \$12,500; ~\$780 per region.



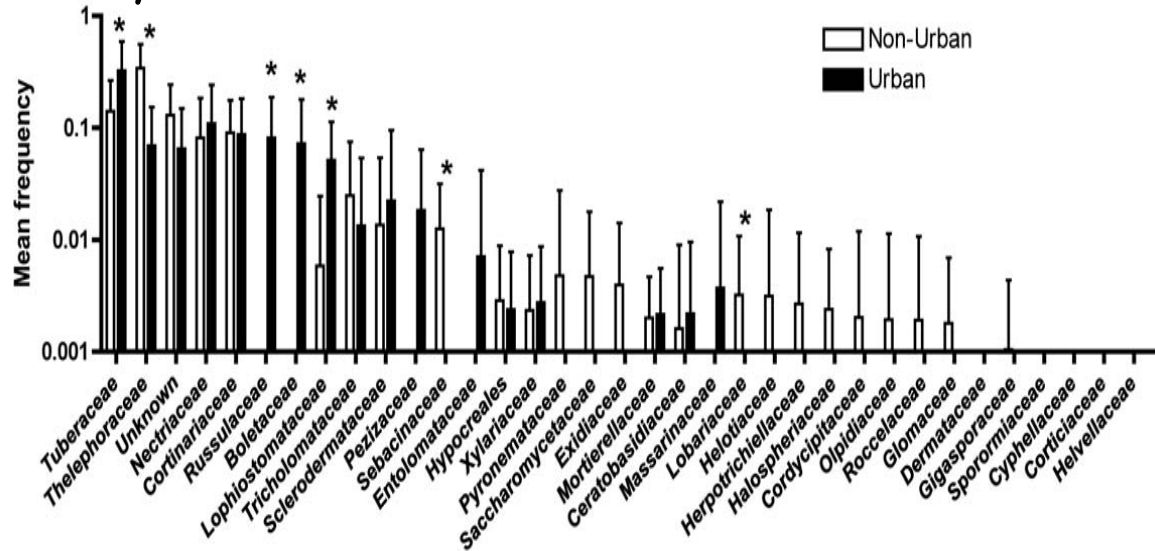
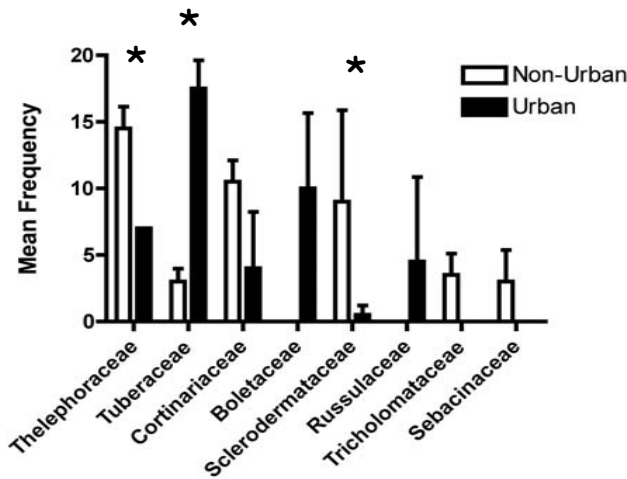
Costs - Urban Ectomycorrhizas

Clone and sequence:

384 reads (4 libraries, 96 reads)
 \$3.65 per read ~ \$1,400
 64 samples: 16 pooled per library

454 sequencing:

~27K reads (3 regions)
 192 samples
 \$780 per region ~ \$2,340



Conclusions:

Thelephoraceae
 Tuberaceae
 Sclerodermataceae
 Boletaceae

Clone

Non-Urban (2x)*
 Urban (6x)*
 Non-Urban (10x)*
 Absent in Non-Urban

454

Non-urban (6x)*
 Urban (6x)*
 Non-Urban (1.5x)^{ns}
 Absent in Non-Urban

Thanks to...

- People

- J. David Mattox - City of Manhattan
- Gary Kilner - KSU
- Chulee Yaege - KSU
- Charles Kramer - KSU
- Nicholas Simpson - KSU
- Ken Jones - UGA

- Funding

- BRIEF - Division of Biology, KSU
- Ecological Genomics Institute, KSU
- National Science Foundation

- Facilities/Infrastructure

- City of Manhattan
- Konza Prairie Biological Station
- Ecological Genomics Institute (KSU)

